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(54) Title: RECOMBINANT 47 AND 31kD COCOA PROTEINS AND PRECURSOR (57) Abstract 47 kD and 31 kD proteins, and their 67 kD expression precursor, believed to be the source of peptide flavour precursors in cocoa (<i>Theobroma cacao</i>) have been identified. Genes coding for them have been probed, identified and sequenced, and recombinant proteins have been synthesised.		

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1 RECOMBINANT 47 AND 31kD COCOA PROTEINS AND PRECURSOR

2

3 This invention relates to proteins and nucleic acids derived from or otherwise
4 related to cocoa.

5

6 The beans of the cocoa plant (*Theobroma cacao*) are the raw material for cocoa,
7 chocolate and natural cocoa and chocolate flavouring. As described by Rohan
8 ("Processing of Raw Cocoa for the Market", FAO/UN (1963)), raw cocoa
9 beans are extracted from the harvested cocoa pod, from which the placenta is
10 normally removed, the beans are then "fermented" for a period of days, during
11 which the beans are killed and a purple pigment is released from the cotyledons.
12 During fermentation "unknown" compounds are formed which on roasting give
13 rise to characteristic cocoa flavour. Rohan suggests that polyphenols and
14 theobromine are implicated in the flavour precursor formation. After
15 fermentation, the beans are dried, during which time the characteristic brown
16 pigment forms, and they are then stored and shipped.

17

18 Biehl *et al*, 1982 investigated proteolysis during anaerobic cocoa seed
19 incubation and identified 26kD and 44kD proteins which accumulated during
20 seed ripening and degraded during germination. Biehl asserted that there were
21 storage proteins and suggested that they may give rise to flavour-specific
22 peptides.

23

24 Fritz *et al*, 1985 identified polypeptides of 20kD and 28kD appearing in the
25 cytoplasmic fraction of cocoa seed extracts at about 100 days after pollination.
26 It appears that the 20kD protein is thought to have glyceryl acyltransferase
27 activity.

28

29 In spite of the uncertainties in the art, as summarised above, proteins apparently
30 responsible for flavour production in cocoa beans have now been identified.
31 Further, it has been discovered that, in spite of Fritz's caution that "cocoa seed

32

33

1 mRNA levels are notably low compared to other plants" (*loc. cit.*), it is possible
2 to apply the techniques of recombinant DNA techniques to the production of
3 such proteins.

4
5 According to a first aspect of the invention, there is provided a 67kD protein of
6 *Theobroma cacao*, or a fragment thereof.

7
8 The 67kD protein appears to be a primary translation product of interest in
9 proteins involved in flavour production in cocoa. The 67kD protein may be
10 processed *in vivo* to form 47kD and 31kD polypeptides.

11
12 According to a second aspect of the invention, there is provided a 47kD protein
13 of *Th. cacao*, or a fragment thereof.

14
15 According to a third aspect of the invention, there is provided a 31kD protein of
16 *Th. cacao* or a fragment thereof.

17
18 The term "fragment" as used herein and as applied to proteins or peptides
19 indicates a sufficient number of amino acid residues are present for the fragment
20 to be useful. Typically, at least four, five, six or even at least 10 or 20 amino
21 acids may be present in a fragment. Useful fragments include those which are
22 the same as or similar or equivalent to those naturally produced during the
23 fermentation phase of cocoa bean processing. It is believed that such fragments
24 take part in Maillard reactions during roasting, to form at least some of the
25 essential flavour components of cocoa.

26
27 Proteins in accordance with the invention may be synthetic; they may be
28 chemically synthesised or, preferably, produced by recombinant DNA
29 techniques. Proteins produced by such techniques can therefore be termed
30 "recombinant proteins". Recombinant proteins may be glycosylated or
31 non-glycosylated: non-glycosylated proteins will result from prokaryotic
32 expression systems.

33

1 *Theobroma cacao* has two primary subspecies, *Th. cacao cacao* and *Th. cacao*
2 *sphaerocarpum*. While proteins in accordance with the invention may be
3 derived from these subspecies, the invention is not limited solely to these
4 subspecies. For example, many cocoa varieties are hybrids between different
5 species; an example of such a hybrid is the trinitario variety.

6
7 The invention also relates to nucleic acid, particularly DNA, coding for the
8 proteins referred to above (whether the primary translation products, the
9 processed proteins or fragments). The invention therefore also provides, in
10 further aspects:

- 11
12 - nucleic acid coding for a 67kD protein of *Th. cacao*, or for a
13 fragment thereof;
14
15 - nucleic acid coding for a 47kD protein of *Th. cacao*, or for a
16 fragment thereof;
17
18 - nucleic acid coding for a 31kD protein of *Th. cacao*, or for a
19 fragment thereof;

20
21 Included in the invention is nucleic acid which is degenerate for the wild type
22 protein and which codes for conservative or other non-deleterious mutants.
23 Nucleic acid which hybridises to the wild type material is also included.

24
25 Nucleic acid within the scope of the invention will generally be recombinant
26 nucleic acid and may be in isolated form. Frequently, nucleic acid in
27 accordance with the invention will be incorporated into a vector (whether an
28 expression vector or otherwise) such as a plasmid. Suitable expression vectors
29 will contain an appropriate promoter, depending on the intended expression
30 host. For yeast, an appropriate promoter is the yeast pyruvate kinase (PK)
31 promoter; for bacteria an appropriate promoter is a strong lambda promoter.

32
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1 Expression may be secreted or non-secreted. Secreted expression is preferred,
2 particularly in eukaryotic expression systems; an appropriate signal sequence
3 may be present for this purpose. Signal sequences derived from the expression
4 host (such as that from the yeast alpha-factor in the case of yeast) may be more
5 appropriate than native cocoa signal sequences.

6
7 The invention further relates to host cells comprising nucleic acid as described
8 above. Genetic manipulation may for preference take place in prokaryotes.
9 Expression will for preference take place in a food-approved host. The yeast
10 *Saccharomyces cerevisiae* is particularly preferred.

11
12 The invention also relates to processes for preparing nucleic acid and protein as
13 described above by nucleic acid replication and expression, respectively.

14
15 cDNA in accordance with the invention may be useful not only for obtaining
16 protein expression but also for Restriction Fragment Length Polymorphism
17 (RFLP) studies. In such studies, detectably labelled cDNA (eg radiolabelled) is
18 prepared. DNA of a cultivar under analysis is then prepared and digested with
19 restriction enzymes. Southern blotting with the labelled cDNA may then enable
20 genetic correlations to be made between cultivars. Phenotypic correlations may
21 then be deduced.

22
23 The invention will now be illustrated by the following non-limiting examples.
24 The examples refer to the accompanying drawings, in which:

25 Figure 1 shows a map of the coding region of the 67kD protein, together with
26 the inter-relationship of plasmids pMS600, pMS700 and pMS800, from which
27 sequence data were obtained;

28
29 Figure 2 shows the complete nucleotide sequence of cDNA coding for the 67kD
30 protein and the deduced amino acid sequence;

31
32 Figure 3 shows the amino acid sequence referred to in Figure 2;
33

1 Figure 4 shows the relationship between the 67kD protein and seed storage
2 proteins from other plants;
3
4 Figure 5 shows a map of plasmid pJLA502;
5
6 Figure 6 shows schematically the formation of plasmid pMS900;
7
8 Figure 7 shows two yeast expression vectors useful in the present invention;
9 vector A is designed for internal expression and vector B is designed for
10 secreted expression;
11
12 Figure 8a shows, in relation to vector A, part of the yeast pyruvate kinase gene
13 showing the vector A cloning site, and the use of *Hin-Nco* linkers to splice in
14 the heterologous gene;
15
16 Figure 8b shows, in relation to vector B, part of the yeast alpha-factor signal
17 sequence showing the vector B cloning site, and the use of *Hin-Nco* linkers to
18 create an in-phase fusion;
19
20 Figure 9a shows how plasmid pMS900 can be manipulated to produce plasmids
21 pMS901, pMS903, pMS907, pMS908, pMS911, pMS912 and pMS914;
22
23 Figure 9b shows how plasmid pMS903 can be manipulated to produce plasmids
24 pMS904, pMS905, pMS906, pMS909 and pMS916;
25
26 Figure 10 shows maps of plasmids pMS908, pMS914, pMS912, pMS906,
27 pMS916 and pMS910;
28
29 Figure 11 shows the construction of a plasmid to express the 67kD protein by
30 means of the AOX promoter on an integrated vector in *Hansenula polymorpha*;
31 and
32
33

Figure 12 shows the construction of a plasmid to express the 67kD protein by means of the AOX promoter in conjunction with the yeast α -factor secretory signal on an integrated vector in *Hansenula polymorpha*.

EXAMPLES

Example 1

Identification of the Major Seed Proteins

It is not practicable to extract proteins directly from cocoa beans due to the high fat and polyphenol contents, and proteins were, therefore, extracted from acetone powders made as follows. Mature beans from cocoa of West African origin (*Theobroma cacao amelonada*) were lyophilised and ground roughly in a pestle and mortar. Lipids were extracted by Soxhlet extraction with diethyl ether for two periods of four hours, the beans being dried and further ground between extractions. Polyphenols and pigments were then removed by several extractions with 80% acetone, 0.1% thioglycolic acid. After extraction the resulting paste was dried under vacuum and ground to a fine powder.

Total proteins were solubilised by grinding the powder with extraction buffer (0.05 M sodium phosphate, pH 7.2; 0.01 M 2-mercaptoethanol; 1% SDS) in a hand-held homogeniser, at 5mg/ml. The suspension was heated at 95°C for 5 minutes, and centrifuged at 18 K for 20 minutes to remove insoluble material. The resulting clear supernatant contained about 1 mg/ml total protein. Electrophoresis of 25 μ l on an SDS-PAGE gel (Laemmli, 1970) gave three major bands, two of which were at 47 kD and 31 kD, comprising over 60% of the total proteins. The 47kD and 31kD proteins are presumed to be the polypeptide subunits of major storage proteins.

1 *Characteristics of the Storage Polypeptides*

2
3 The solubility characteristics of the 47 kD and 31 kD polypeptides were roughly
4 defined by one or two quick experiments. Dialysis of the polypeptide solution
5 against SDS-free extraction buffer rendered the 47 kD and 31 kD polypeptides
6 insoluble, as judged by their ability to pass through a 0.22 micron membrane.
7 Fast Protein Liquid Chromatography (FPLC) analysis also showed that the 47
8 kD and 31 kD polypeptides were highly associated after extraction with
9 McIlvaines buffer pH 6.8 (0.2 M disodium hydrogen phosphate titrated with
10 0.1 M citric acid). The 47 kD and 31 kD polypeptides are globulins on the
11 basis on their solubility.

12

13 *Purification of the 47 kD and 31 kD polypeptides*

14

15 The 47 kD and 31 kD polypeptides were purified by two rounds of gel filtration
16 on a SUPEROSE-12 column of the PHARMACIA Fast Protein Liquid
17 Chromatography system (FPLC), or by electroelution of bands after preparative
18 electrophoresis. (The words SUPEROSE and PHARMACIA are trade marks.)
19 Concentrated protein extracts were made from 50 mg acetone powder per ml of
20 extraction buffer, and 1-2 ml loaded onto 2 mm thick SDS-PAGE gels poured
21 without a comb. After electrophoresis the gel was surface stained in aqueous
22 Coomassie Blue, and the 47 kD and 31 kD bands cut out with a scalpel. Gel
23 slices were electroeluted into dialysis bags in electrophoresis running buffer at
24 15 V for 24 hours, and the dialysate dialysed further against 0.1% SDS.
25 Samples could be concentrated by lyophilisation.

26

27 Example 2

28

29 *Amino-acid Sequence Data from Proteins*

30

31 Protein samples (about 10 μ g) were subjected to conventional N-terminal
32 amino-acid sequencing. The 47 kD and 31 kD polypeptides were N-terminally
33 blocked, so cyanogen bromide peptides of the 47 kD and 31 kD peptides were

1 prepared, and some amino-acid sequence was derived from these. Cyanogen
2 bromide cleaves polypeptide chains at methionine residues, and thus cleaved the
3 47 kD and 31 kD polypeptides gave rise to 24 kD and 17 kD peptides. In
4 addition the 47 kD polypeptide gave a 20 kD peptide. The 24 kD and 17 kD
5 peptides had the same 9 N-terminal amino-acid residues. This fact, combined
6 with the obvious one that the 31 kD could not contain both peptides
7 consecutively, suggested that the 24 kD peptide arose for a partial digest, where
8 full digestion would yield the 17 kD peptide. The other striking conclusion is
9 that the 47 kD and 31 kD proteins are related, and the 31 kD could be a further
10 processed form of the 47 kD. The 9 amino-acid sequence was used to construct
11 an oligonucleotide probe for the 47 kD/31 kD gene(s).

12

13 Example 3

14

15 *Raising Antibodies to the 47 kD and 31 kD Polypeptides*

16

17 Polyclonal antibodies were prepared using the methodology of Catty and
18 Raykundalia (1988). The serum was aliquoted into 1 ml fractions and stored at
19 -20°C.

20

21 *Characterising Antibodies to the 47 kD and 31 kD Polypeptides*

22

23 Serum was immediately characterised using the Ochterloney double-diffusion
24 technique, whereby antigen and antibody are allowed to diffuse towards one
25 another from wells cut in agarose in borate-saline buffer. Precipitin lines are
26 formed where the two interact if the antibody 'recognises' the antigen. This test
27 showed that antibodies to both antigens had been formed, and furthermore that
28 extensive cross-reaction took place between the 47 kD and 31 kD polypeptides
29 and their respective antibodies. This is further indication that the 47 kD and 31
30 kD polypeptides are closely related, as suggested by their cyanogen bromide
31 cleavage patterns.

32

33

1 The gamma-globulin fraction of the serum was partially purified by
2 precipitation with 50% ammonium sulphate, solubilisation in
3 phosphate-buffered saline (PBS) and chromatography on a DE 52 cellulose
4 ion-exchange column as described by Hill, 1984. Fractions containing
5 gamma-globulin were monitored at 280 nm (OD_{280} of 1.4 is equivalent to 1
6 mg/ml gamma-globulin) and stored at -20°C .
7 The effective titre of the antibodies was measured using an enzyme-linked
8 immunosorbant assay (ELISA). The wells of a polystyrene microtitre plate
9 were coated with antigen (10-1000 ng) overnight at 4°C in carbonate coating
10 buffer. Wells were washed in PBS-Tween and the test gamma globulin added at
11 concentrations of 10, 1 and $0.1\text{ }\mu\text{g/ml}$ (approximately 1:100, 1:1000 and
12 1:10,000 dilutions). The diluent was PBS-Tween containing 2% polyvinyl
13 pyrrolidone (PVP) and 0.2% BSA. Controls were preimmune serum from the
14 same animal. Binding took place at 37°C for 3-4 hours. The wells were
15 washed as above and secondary antibody (goat anti-rabbit IgG conjugated to
16 alkaline phosphatase) added at a concentration of $1\text{ }\mu\text{g/ml}$, using the same
17 conditions as the primary antibody. The wells are again washed, and alkaline
18 phosphatase substrate (p-nitrophenyl phosphate; 0.6 mg/ml in diethanol-amine
19 buffer pH 9.8) added. The yellow colour, indicating a positive reaction, was
20 allowed to develop for 30 minutes and the reaction stopped with 3M NaOH.
21 The colour is quantified at 405 nm. More detail of this method is given in Hill,
22 1984. The method confirmed that the antibodies all had a high titre and could
23 be used at $1\text{ }\mu\text{g/ml}$ concentration.

24

25 Example 4

26

27 *Isolation of Total RNA from Immature Cocoa Beans*

28

29 The starting material for RNA which should contain a high proportion of
30 mRNA specific for the storage proteins was immature cocoa beans, at about 130
31 days after pollination. Previous work had suggested that synthesis of storage
32 proteins was approaching its height by this date (Biehl *et al.*, 1982). The beans
33 are roughly corrugated and pale pinkish-purple at this age.

1
2 The initial requirement of the total RNA preparation from cocoa beans was that
3 it should be free from contaminants, as judged by the UV spectrum, particularly
4 in the far UV, where a deep trough at 230 nm (260 nm : 230 nm ratio is
5 approximately 2.0) is highly diagnostic of clean RNA, and is intact, as judged
6 by agarose gel electrophoresis of heat-denatured samples, which should show
7 clear rRNA bands. A prerequisite for obtaining intact RNA is scrupulous
8 cleanliness and rigorous precautions against RNases, which are ubiquitous and
9 extremely stable enzymes. Glassware is customarily baked at high
10 temperatures, and solutions and apparatus treated with the RNase inhibitor
11 diethyl pyrocarbonate (DEPC, 0.1%) before autoclaving.
12

13 The most routine method for extraction of plant (and animal) RNA is extraction
14 of the proteins with phenol/chloroform in the presence of SDS to disrupt
15 protein-nucleic acid complexes, and inhibit the RNases which are abundant in
16 plant material. Following phenol extraction the RNA is pelleted on a caesium
17 chloride gradient before or after ethanol precipitation. This method produced
18 more or less intact RNA, but it was heavily contaminated with dark brown
19 pigment, probably oxidised polyphenols and tannins, which always co-purified
20 with the RNA. High levels of polyphenols are a major problem in *Theobroma*
21 tissues.
22

23 A method was therefore adopted which avoided the use of phenol, and instead
24 used the method of Hall *et al.* (1978) which involves breaking the tissue in hot
25 SDS-borate buffer, digesting the proteins with proteinase K, and specifically
26 precipitating the RNA with LiCl. This method gave high yields of reasonably
27 clean, intact RNA. Contaminants continued to be a problem and the method
28 was modified by introducing repeated LiCl precipitation steps, the precipitate
29 being dissolved in water and clarified by microcentrifugation after each step.
30 This resulted in RNA preparations with ideal spectra, which performed well in
31 subsequent functional tests such as *in vitro* translation.
32
33

1 *Preparation of mRNA From Total RNA*

2

3 The mRNA fraction was separated from total RNA by affinity chromatography
4 on a small (1 ml) oligo-dT column, the mRNA binding to the column by its
5 poly A tail. The RNA (1-2 mg) was denatured by heating at 65°C and applied
6 to the column in a high salt buffer. Poly A+ was eluted with low salt buffer,
7 and collected by ethanol precipitation. The method is essentially that of Aviv
8 and Leder (1972), modified by Maniatis *et al* (1982). From 1 mg of total
9 RNA, approximately 10-20 µg polyA+ RNA was obtained (1-2%).

10

11 *In vitro Translation of mRNA*

12

13 The ability of mRNA to support *in vitro* translation is a good indication of its
14 cleanliness and intactness. Only mRNAs with an intact polyA tail (3' end) will
15 be selected by the oligo-dT column, and only mRNAs which also have an intact
16 5' end (translational start) will translate efficiently. *In vitro* translation was
17 carried out using RNA-depleted wheat-germ lysate (Amersham International),
18 the *de novo* protein synthesis being monitored by the incorporation of [³⁵
19 S]-methionine (Roberts and Paterson, 1973). Initially the rate of *de novo*
20 synthesis was measured by the incorporation of [³⁵ S]-methionine into
21 TCA-precipitable material trapped on glass fibre filters (GFC, Whatman). The
22 actual products of translation were investigated by running on SDS-PAGE,
23 soaking the gel in fluor. drying the gel and autoradiography. The mRNA
24 preparations translated efficiently and the products covered a wide range of
25 molecular weights, showing that intact mRNAs for even the largest proteins had
26 been obtained. None of the major translation products corresponded in size to
27 the 47kD or 31kD storage polypeptides identified in mature beans, and it was
28 apparent that considerable processing of the nascent polypeptides must occur to
29 give the mature forms.

30

31

32

33

1 Example 5

2

3 *Identification of Precursor to the 47 kD and 31 kD Polypeptides by*
4 *Immunoprecipitation*

5

6 Because the 47 kD and 31 kD storage polypeptides were not apparent amongst
7 the translation products of mRNA from developing cocoa beans, the technique
8 of immunoprecipitation, with specific antibodies raised to the storage
9 polypeptides, was used to identify the precursors from the translation mixture.
10 This was done for two reasons: first to confirm that the appropriate mRNA was
11 present before cloning, and second to gain information on the expected size of
12 the encoding genes.

13

14 Immunoprecipitation was by the method of Cuming *et al*, 1986. [³⁵S]-labelled
15 *in vitro* translation products were dissociated in SDS, and allowed to bind with
16 specific antibody in PBS plus 1% BSA. The antibody-antigen mixture was then
17 mixed with protein A-SEPHAROSE and incubated on ice to allow the IgG to
18 bind to protein A. The slurry was poured into a disposable 1 ml syringe, and
19 unbound proteins removed by washing with PBS +1% NONIDET P-40. The
20 bound antibody was eluted with 1M acetic acid and the proteins precipitated
21 with TCA. The antibody-antigen complex was dissociated in SDS, and subject
22 to SDS-PAGE and fluorography, which reveals which labelled antigens have
23 bound to the specific antibodies.

24

25 The results showed that the anti-47 kD and anti-31 kD antibodies both
26 precipitated a 67 kD precursor. The precursor size corresponded to a major
27 band on the *in vitro* translation products. The results with the 47 kD and 31 kD
28 antibodies confirmed that the polypeptides are derived from a single precursor,
29 or at least precursors of the same size. The large size of the precursor
30 suggested that size-selection at mRNA or cDNA level may be necessary to
31 obtain clones.

32

33

1 Example 6

2

3 *cDNA Synthesis From the mRNA Preparations*

4

5 cDNA synthesis was carried out using a kit from Amersham International. The
6 first strand of the cDNA is synthesised by the enzyme reverse transcriptase,
7 using the four nucleotide bases found in DNA (dATP, dTTP, dGTP, dCTP) and
8 an oligo-dT primer. The second strand synthesis was by the method of Gubler
9 and Hoffman (1983), whereby the RNA strand is nicked in many positions by
10 RNase H, and the remaining fragments used to prime the replacement synthesis
11 of a new DNA strand directed by the enzyme *E. coli* DNA polymerase I. Any
12 3' overhanging ends of DNA are filled in using the enzyme T4 polymerase.
13 The whole process was monitored by adding a small proportion of [³²P]-dCTP
14 into the initial nucleotide mixture, and measuring the percentage incorporation
15 of label into DNA. Assuming that cold nucleotides are incorporated at the same
16 rate, and that the four bases are incorporated equally, an estimate of the
17 synthesis of cDNA can be obtained. From 1 µg of mRNA approximately 140
18 ng of cDNA was synthesised. The products were analysed on an alkaline 1.4%
19 agarose gel as described in the Amersham methods. Globin cDNA, synthesised
20 as a control with the kit, was run on the same gel, which was dried down and
21 autoradiographed. The cocoa cDNA had a range of molecular weights, with a
22 substantial amount larger than the 600 bp of the globin cDNA.

23

24 Example 7

25

26 *Cloning of cDNA into a Plasmid Vector by Homopolymer Tailing*

27

28 The method of cloning cDNA into a plasmid vector was to 3' tail the cDNA
29 with dC residues using the enzyme terminal transferase (Boehringer Corporation
30 Ltd), and anneal into a *Pst*I-cut and 5' tailed plasmid (Maniatis *et al*, 1982
31 Eschenfeldt *et al*, 1987). The optimum length for the dC tail is 12-20 residues.
32 The tailing reaction (conditions as described by the manufacturers) was tested

33

1 with a 1.5 kb blunt-ended restriction fragment, taking samples at intervals, and
2 monitoring the incorporation of a small amount of [³²P]-dCTP. A sample of
3 cDNA (70 ng) was then tailed using the predetermined conditions.

4
5 A dG-tailed plasmid vector (3'-oligo(dG)-tailed pUC9) was purchased from
6 Pharmacia. 15 ng vector was annealed with 0.5 - 5 ng of cDNA at 58°C for 2
7 hours in annealing buffer: 5mM Tris-HCl pH 7.6; 1mM EDTA, 75 mM NaCl
8 in a total volume of 50 µl. The annealed mixture was transformed into *E. coli*
9 RRI (Bethesda Research Laboratories), transformants being selected on L-agar
10 + 100 µg/ml ampicillin. Approximately 200 transformants per ng of cDNA
11 were obtained. Transformants were stored by growing in 100 µl L-broth in the
12 wells of microtitre plates, adding 100 µl 80% glycerol, and storing at -20°C.

13
14 Some of the dC tailed cDNA was size selected by electrophoresing on a 0.8%
15 agarose gel, cutting slits in the gel at positions corresponding to 0.5, 1.0 and
16 1.5 kb, inserting DE81 paper and continuing electrophoresis until the cDNA
17 had run onto the DE81 paper. The DNA was then eluted from the paper with
18 high salt buffer, according to the method of Dretzen *et al* (1981).

19 20 21 Example 8

22 23 *Construction of Oligonucleotide Probes for the 47/31 kD Gene*

24
25 The amino-acid sequence obtained from a cyanogen bromide peptide common to
26 the 47 kD and 31 kD polypeptides is as follows:

27
28 Met-Phe-Glu-Ala-Asn-Pro-Asn-Thr-Phe
29

30 and the least redundant probe of 17 residues (a mixture of 32) is shown below:
31
32
33

1
2 Met-Phe-Glu-Ala-Asn-Pro
3 5' ATG TTT GAA GCT AAT CC 3'
4 C G C C
5 A
6 G

7 The actual probe was made anti-sense so that it could also be used to probe
8 mRNA. Probe synthesis was carried out using an Applied Biosystems
9 apparatus.

10
11
12 Example 9

13
14 *Use of Oligonucleotides to Probe cDNA Library*

15
16 The oligonucleotide probes were 5' end-labelled with gamma-[³²P] dATP and
17 the enzyme polynucleotide kinase (Amersham International). The method was
18 essentially that of Woods (1982, 1984), except that a smaller amount of isotope
19 (15 µCi) was used to label about 40 ng probe, in 10 mM MgCl₂, 100 mM
20 Tris-HCl, pH 7.6; 20 mM 2-mercaptoethanol.

21
22 The cDNA library was grown on GeneScreen (New England Nuclear) nylon
23 membranes placed on the surface of L-agar + 100 µg/ml ampicillin plates. (The
24 word GeneScreen is a trade mark.) Colonies were transferred from microtitre
25 plates to the membranes using a 6 x 8 multi-pronged device, designed to fit into
26 the wells of half the microtitre plate. Colonies were grown overnight at 37°C,
27 lysed in sodium hydroxide and bound to membranes as described by Woods
28 (1982, 1984). After drying the membranes were washed extensively in 3 x
29 SSC/0.1% SDS at 65°C, and hybridised to the labelled probe, using a HYBAID
30 apparatus from Hybaid Ltd, PO Box 82, Twickenham, Middlesex. (The word
31 HYBAID is a trade mark.) Conditions for hybridisation were as described by
32 Mason & Williams (1985), a T_d being calculated for each oligonucleotide
33 according to the formula:

$T_d = 4^{\circ}\text{C}$ per GC base pair + 2°C per AT base pair.
At mixed positions the lowest value is taken.

Hybridisation was carried out at $T_d - 5^{\circ}\text{C}$. Washing was in 6 x SSC, 0.1% SDS initially at room temperature in the HYBAID apparatus, then at the hybridisation temperature ($T_d - 5^{\circ}\text{C}$) for some hours, and finally at T_d for exactly 2 minutes. Membranes were autoradiographed onto FUJI X-ray film, with intensifying screens at -70°C . (The word FUJI is a trade mark.) After 24 - 48 hours positive colonies stood out as intense spots against a low background.

Example 10

Analysis of Positive Clones for the 47 kD/31 kD Polypeptide

Only one positive clone, pMS600, was obtained. This released two *Pst*I fragments on digestion, of total length 1.3 kb, insufficient to encode the 67 kD precursor. The total insert was removed from the vector on a *Hind*III-*Eco*RI fragment, nick-translated and used to probe the cDNA library, picking up a further two positive clones, pMS700 and pMS800. Restriction mapping of all three inserts suggested an overlapping map covering nearly 2.0 kb, sufficient to encode the 67 kD precursor (see Figure 1).

Example 11

Sequencing the Cloned Inserts

The sequencing strategy was to clone the inserts, and where appropriate subclones thereof, into the multiple cloning site of the plasmids pTZ18R/pTZ19R (Pharmacia). These plasmids are based on the better-known vectors pUC18/19 (Norrandar *et al.* 1983), but contain a single-stranded origin of replication from the filamentous phage f1. When superinfected with phages in the same group, the plasmid is induced to undergo single-stranded

1 replication, and the single-strands are packaged as phages extruded into the
2 medium. DNA can be prepared from these 'phages' using established methods
3 for M13 phages (Miller, 1987), and used for sequencing by the method of
4 Sanger (1977) using the reverse sequencing primer. The superinfecting phage
5 used is a derivative of M13 termed M13K07, which replicates poorly and so
6 does not compete well with the plasmid, and contains a selectable
7 kanamycin-resistance marker. Detailed methods for preparing single-strands
8 from the pTZ plasmids and helper phages are supplied by Pharmacia. DNA
9 sequence was compiled and analysed using the Staden package of programs
10 (Staden, 1986), on a PRIME 9955 computer. (The word PRIME is a trade
11 mark.)
12

13 Example 12

14 *Features of the 47 kD/31 kD cDNA and Deduced Amino-acid Sequence of the 67* 15 *kD Precursor*

16
17
18 DNA sequencing of the three positive clones, pMS600, pMS700, pMS800,
19 confirmed the overlap presumed in Figure 1. No sequence differences were
20 found in the overlapping regions (about 300 bp altogether), suggesting that the
21 three cDNAs were derived from the same gene. The sequence of the combined
22 cDNAs comprising 1818 bases is shown in Figure 2. The first ATG codon is
23 found at position 14, and is followed by an open reading frame of 566 codons.
24 There is a 104-base 3' untranslated region containing a polyadenylation signal at
25 position 1764. The oligonucleotide probe sequence is found at position 569.
26

27 The open reading frame translates to give a polypeptide of 566 amino-acids
28 (Figure 2), and a molecular weight of 65612, which is reasonably close to the
29 67 kD measured on SDS-PAGE gels. The N-terminal residues are clearly
30 hydrophobic and look like a characteristic signal sequence. Applying the rules
31 of Von Heije (1983), which predict cleavage sites for signal sequences, suggests
32 a cleavage point between amino-acids 20 and 21 (see Figure 3). The region
33 following this is highly hydrophobic and contains four Cys-X-X-X-Cys motifs.

1 The N-terminus of the mature protein has been roughly identified as the
2 glutamate (E) residue at 135 (Figure 3), on the basis of some tentative
3 N-terminal sequence (EEPGSQFANPAYHF). This N-terminus would give a
4 mature protein of 49068 kD, in rough agreement with that observed. There
5 appears to be no glycosylation sites (Asn-X-Ser/Thr) in the mature protein of
6 the sequence.

7
8 *Homologies Between the 67 kD Precursor and Other Known Proteins*
9

10 Searches through the PIR database, and through the literature, revealed close
11 homologies between the 67 kD polypeptide and a class of seed storage proteins
12 termed vicilins, one of two major classes of globulins found in seed (Borrito
13 and Dure, 1987). Alignments between the 67 kD polypeptide and vicilins from
14 cotton (*Gossypium hirsutum*, Ghi), soybean (*Glycine max*, Gma), pea (*Pisum*
15 *sativum*, Psa-c is convicilin, Psa-v is vicilin) and bean (*Phaseolus vulgaris*, Pvu)
16 are shown in Figure 4 (Bown *et al*, 1988; Chlan *et al*, 1986; Doyle *et al*, 1986;
17 Lycett *et al*, 1983). Identical residues are boxed.

18
19 All the vicilins have a mature molecular weight of around 47 kD, with the
20 exception of soybean conglycinin alpha and alpha1 subunits, which are 72 kD
21 and 76 kD respectively, and pea convicilin with a mature molecular weight of
22 64kD. The pea and bean subunits (2 sub classes each) are synthesised as small
23 precursors, around 50 kD. The most striking homology with the 67 kD is the
24 cotton vicilin (Chlan *et al*, 1986). Cotton is also the most closely related to
25 cocoa: both are members of the order Malvales. Interestingly cotton also has a
26 large precursor, of 69 kD, comprising a short signal sequence, a large
27 hydrophilic domain containing six Cys-X-X-X-Cys motifs, and a mature
28 domain. It may therefore be possible to synthesise the corresponding cotton
29 protein, by techniques analogous to those disclosed in this application and to use
30 the cotton protein, or fragments of it, in the preparation of flavour components
31 analogous to cocoa flavour components.
32
33

1 Example 13

2

3 *Expression of the 67 kD Polypeptide in E. coli*

4

5 Before the 67 kD coding region could be inserted into a expression vector the
6 overlapping fragments from the three separate positive clones had to be spliced
7 into a continuous DNA segment. The method of splicing is illustrated in Figure
8 6: a *HindIII*-*BglIII* fragment from pMS600, a *BglIII*-*EcoRI* fragment from
9 pMS700 and an *EcoRI*-*SalI* fragment from pMS800 were ligated into pTZ19R
10 cut with *HindIII* and *SalI*. The resulting plasmid, containing the entire 67 kD
11 cDNA, was termed pMS900.

12

13 An *NcoI* site was introduced at the ATG start codon, using the mutagenic
14 primer:

15

16 5' TAG CAA CCA TGG TGA TCA 3'.

17

18 *In vitro* mutagenesis was carried out using a kit marketed by Amersham
19 International, which used the method of Eckstein and co-workers (Taylor *et al*,
20 1985). After annealing the mutagenic primer to single-stranded DNA the
21 second strand synthesis incorporates alpha-thio-dCTP in place of dCTP. After
22 extension and ligation to form closed circles, the plasmid is digested with *NciI*,
23 an enzyme which cannot nick DNA containing thio-dC. Thus only the original
24 strand is nicked, and subsequently digested with exonuclease III. The original
25 strand is then resynthesised, primed by the remaining DNA fragments and
26 complementing the mutated position in the original strand. Plasmids are then
27 transformed into *E. coli* and checked by plasmid mini preparations.

28

29 The 67 kD cDNA was then cloned into the *E. coli* expression plasmid, pJLA502
30 (Figure 5), on an *NcoI* - *SalI* fragment (pMS902).

31

32

33

pJLA502 (Schauder *et al*, 1987) is marketed by Medac GmbH, Postfach 303629, D-7000, Hamburg 36 and contains the strong lambda promoters, P_L and P_R , and the leader sequence and ribosome binding site of the very efficiently translated *E. coli* gene, *atpE*. It also contains a temperature-sensitive cI repressor, and so expression is repressed at 30°C and activated at 42°C. The vector has an *NcoI* site (containing an ATG codon: CCATGG) correctly placed with respect to the ribosome binding site, and foreign coding sequences must be spliced in at this point.

The expression vector was transformed into *E. coli* UT580. The transformed strain was grown in L-broth + ampicillin (100 µg/ml) at 30°C until log phase ($OD_{610} = 0.5$) and the temperature was then shifted to 42°C and samples taken at intervals. Samples were dissociated by boiling in SDS loading buffer, and run on SDS-PAGE gels. The proteins were electroblotted onto nitrocellulose membranes (Towbin *et al*, 1979) and Western blotting carried out using the anti-21 kD antibody prepared in Example 3 above (at 2 µg/ml) and as a secondary antibody, goat anti-rabbit-IgG conjugated to alkaline phosphatase (Scott *et al*, 1988).

A specific band at 67 kD was produced by pMS902, showing that a functional gene was present.

Example 14

Expression of the 67 kD Polypeptide in Yeast

Two yeast expression vectors were used, both based on a yeast-*E. coli* shuttle vector containing yeast and *E. coli* origins of replication, and suitable selectable markers (ampicillin-resistance for *E. coli* and leucine auxotrophy for yeast). Both vectors contain the yeast pyruvate kinase (PK) promoter and leader sequence and have a *HindIII* cloning site downstream of the promoter. One vector, A (YVA), is designed for internal expression, and the other, B (YVB), for secreted expression, having a portion of the signal sequence of the yeast

1 mating alpha-factor downstream of the promoter, with a *HindIII* site within it to
2 create fusion proteins with incoming coding sequences. The vectors are
3 illustrated in Figure 7.

4
5 To use the vectors effectively it is desirable to introduce the foreign coding
6 region such that for vector A, the region from the *HindIII* cloning site to the
7 ATG start is the same as the yeast PK gene, and for vector B, the remainder of
8 the alpha-factor signal, including the lysine at the cleavage point. In practice
9 this situation was achieved by synthesising two sets of *HindIII* - *NcoI* linkers to
10 breach the gap between the *HindIII* cloning site in the vector and the *NcoI* at the
11 ATG start of the coding sequence. This is illustrated in Figure 8.

12
13 In order to use the yeast vector B, the hydrophobic signal sequence must first be
14 deleted from the 67 kD cDNA. Although direct evidence of the location of the
15 natural cleavage site was lacking, the algorithm of Von Heije predicts a site
16 between amino-acids 20 (alanine) and 21 (leucine). However it was decided to
17 remove amino-acids 2-19 by deletion, so that the useful *NcoI* site at the
18 translation start would be maintained.

19

20

21 For ease of construction of the yeast vectors, the strategy was to first clone the
22 *HindIII* - *NcoI* linkers into the appropriate pTZ plasmids, and then to clone the
23 linkers plus coding region into the yeast vectors on *HindIII* - *BamHI* fragments.
24 However the coding region contains an internal *BamHI* which must be removed
25 by in vitro mutagenesis, giving a new plasmid pMS903. The signal sequence
26 was deleted from pMS903 using the mutagenic primer

27

28 5' AGCATAGCAACCATGGTTGCTTTGTTCT 3'

29

30 to give pMS904. The appropriate *HindIII* - *NcoI* linkers were then cloned into
31 pMS903 and pMS904 to give pMS907 and pMS905 respectively, and the
32 *HindIII* - *BamHI* fragments (linkers + coding region) subcloned from these

33

1 intermediate plasmids into YVA and YVB respectively to give the yeast
2 expression plasmids pMS908 and pMS906. A diagrammatic scheme for these
3 and other constructs is given in Figure 9.

4
5 Because the mature cocoa protein appears to lack the N-terminal hydrophilic
6 domain, as described in Example 12, expression vectors have also been
7 designed to express the mature protein directly. Yeast is unlikely to have the
8 same processing enzymes as cocoa and optimum expression may be obtained for
9 a protein as close as possible to that found naturally in cocoa. Hence the DNA
10 encoding the hydrophilic domain (amino acids 20-134) was deleted from the
11 intermediate plasmids pMS907 and pMS905 to give plasmids pMS911 and
12 pMS909 respectively, and the *Hind*III - *Bam*HI fragments for these were cloned
13 into YVA and YVB to give the expression plasmids pMS912 and pMS910
14 (Figure 9).

15
16 A further modification was introduced by constructing expression in which the
17 plant terminator had been removed and replaced with the yeast ADH terminator
18 (present in YVA and YVB). The plant signal was removed by cutting the
19 intermediate plasmids pMS907 and pMS905 at the *Pvu*II site immediately
20 downstream of the coding region, at position 1716 in Figure 2. *Hind*III linkers
21 were added and the entire coding region cloned into the yeast expression vectors
22 on *Hind*III - *Hind*III fragments giving expression plasmids pMS914 (YVA) and
23 pMS916 (YVB) (Figure 9). A summary of the constructs made is given in
24 Figure 10.

25
26 The yeast expression plasmids were transferred into yeast spheroplasts using the
27 method of Johnston (1988). The transformation host was the LEU⁻ strain
28 AH22, and transformants were selected on leucine-minus minimal medium.
29 LEU⁺ transformants were streaked to single colonies, which were grown in 50
30 ml YEPD medium (Johnston, 1988) at 28 °C for testing the extent and
31 distribution of foreign protein. Cells were harvested from cultures in
32 preweighed tubes in a bench-top centrifuge, and washed in 10 ml lysis buffer
33 (200 mM Tris, pH 8.1: 10% glycerol). The cell medium was reserved and

1 concentrated 10-25 x in an AMICON mini concentrator. (The word AMICON
2 is a trade mark.) The washed cells were weighed and resuspended in lysis
3 buffer plus protease inhibitors (1 mM phenyl methyl sulphonyl fluoride
4 (PMSF); 1 µg/ml aprotinin; 0.5 µg/ml leupeptin) at a concentration of 1 g/ml.
5 1 volume acid-washed glass-beads was added and the cells broken by vortexing
6 for 8 minutes in total, in 1 minute bursts, with 1 minute intervals on ice. After
7 checking under the microscope for cell breakage, the mixture was centrifuged at
8 7000 rpm for 3 minutes to pellet the glass beads. The supernatant was removed
9 to a pre-chilled centrifuge tube, and centrifuged for 1 hour at 20,000 rpm.
10 (Small samples can be centrifuged in a microcentrifuge in the cold.) The
11 supernatant constitutes the soluble fraction. The pellet was resuspended in 1 ml
12 lysis buffer plus 10% SDS and 1% mercaptoethanol and heated at 90°C for 10
13 minutes. After centrifuging for 15 minutes in a microcentrifuge the supernatant
14 constitutes the particulate fraction.

15

16 Samples of each fraction and the concentrated medium were examined by
17 Western blotting. Considering first the plasmids designed for internal
18 expression in YVA, pMS908 produced immunoreactive proteins at 67 kD and
19 16 kD within the cells only. There was no evidence of the 67 kD protein being
20 secreted under the influence of its own signal sequence. The smaller protein is
21 presumed to be a degradation product. A similar result, but with improved
22 expression, was obtained with pMS914, in which the plant terminator is
23 replaced by a yeast terminator. However in pMS912, in which the coding
24 region for the hydrophilic domain has been deleted, no synthesis of
25 immunoreactive protein occurred.

26

27 For industrial production of heterologous proteins in yeast a secreted mode is
28 preferable because yeast cells are very difficult to disrupt, and downstream
29 processing from total cell protein is not easy. The results from the vectors
30 constructed for secreted expressed were rather complicated. From the simplest
31 construct, pMS906, in which the yeast α -factor signal sequence replaces the
32 plant protein's own signal, immunoreactive proteins of approximately 47 kD, 28
33 kD and 18-20 kD were obtained and secreted into the medium. At first sight

1 this is surprising because the coding region introduced should synthesise a 67
2 kD protein. However the most likely explanation is that the yeast's KEX2
3 protease, that recognises and cleaves the α -factor signal at a Lys-Arg site is also
4 cleaving the 67 kD protein at Lys-Arg dipeptides at positions 148 and 313 in
5 the amino-acid sequence. The calculated protein fragment sizes resulting from
6 cleavage at these positions are 47179 Daltons, 28344 Daltons and 18835
7 Daltons, very close to the observed sizes.

8
9 When the plant terminator is replaced with a yeast terminator in pMS916 no
10 expression is obtained in either cells or medium; it is possible that a mutation
11 has been inadvertently introduced. From the construct pMS910, in which the
12 hydrophilic domain has been deleted the main antigenic products were 28 kD
13 and 18-20 kD, again secreted into the medium. It is presumed that the *de novo*
14 47 kD product is immediately cleaved at the KEX2 site at position 313.

15
16 In summary, four of the six expression vectors constructed direct the synthesis
17 of proteins cross-reacting with anti-47 kD antibodies. Two of the constructs
18 secrete proteins into the medium.

19 20 Example 15

21 22 *Construction of Vectors Designed to Express the 67 kD Protein in Hansenula* 23 *polymorpha*

24
25 The methylotropic yeast *Hansenula polymorpha* offers a number of advantages
26 over *Saccharomyces cerevisiae* as a host for the expression of heterologous
27 proteins (EP-A-0173378 and Sudbery *et al.*, 1988). The yeast will grow on
28 methanol as sole carbon source, and under these conditions the enzyme
29 methanol oxidase (MOX) can represent up to 40% of the total cell protein.
30 Thus the MOX promoter is a very powerful one that can be used in a vector to
31 drive the synthesis of heterologous proteins, and it is effective even as a single
32 copy. This gives the potential to use stable integrated vectors. *Hansenula* can
33 also grow on rich carbon sources such as glucose, in which case the MOX

1 promoter is completely repressed. This means that cells containing the
2 heterologous gene can be grown to a high density on glucose, and induced to
3 produce the foreign protein by allowing the glucose to run out and adding
4 methanol.

5
6 A plasmid, pHGL1, containing the MOX promoter and terminator, and a
7 cassette containing the yeast α -factor secretory signal sequence, were prepared.
8 The 67 kD coding region was cloned into pHGL1 on a *Bam*HI - *Bam*HI
9 fragment, replacing the *Bgl*II fragment which contains the 3' end of the MOX
10 coding region. The whole promoter - gene - terminator region can then be
11 transferred to YEp13 on a *Bam*HI - *Bam*HI fragment to give the expression
12 plasmid pMS922. The details of the construction are illustrated in Figure 11.
13 An analogous expression plasmid, pMS925, has been constructed with the yeast
14 α -factor spliced onto the 67 kD coding region, replacing the natural plant
15 signal. The *Bam*HI - *Hind*III cassette containing the α -factor was ligated to the
16 *Hind*III - *Bam*HI fragment used to introduce the 67 kD coding region into YVB.
17 The α -factor plus coding region was then cloned with pHGL1 on a *Bam*HI -
18 *Bam*HI fragment, and transferred into YEP13 as before. Details are shown in
19 Figure 12.

20
21 Both constructs have been transformed into *Hansenula* and grown under
22 inducing conditions with 0.5% or 1% methanol. Both constructs directed the
23 production of immunoreactive protein within the cells, and pMS925 secreted the
24 protein into the medium under the influence of the α -factor signal sequence.

25

26 *E. coli* Strains

27

28 RR1 $F^+v_B^-M_B$ *ara*-14 *pro*A2 *leu*B6 *lac*Y1 *gal*K2 *vps*L20 (*str*^r)
29 *xyl*-5 *mtl*-1 *sup*E44 -

30

31 CAG629 *lac*_{am} *rvp*_{am} *pho*_{am} *htpR*_{am} *mal* *rps*L *lon* *sup*C_{ts}

32

33

1 UT580 (lac-pro) supE thi hsdD5 / F' tra D36 proA⁺B⁺ lacI^q lacZ
2 M15
3
4
5

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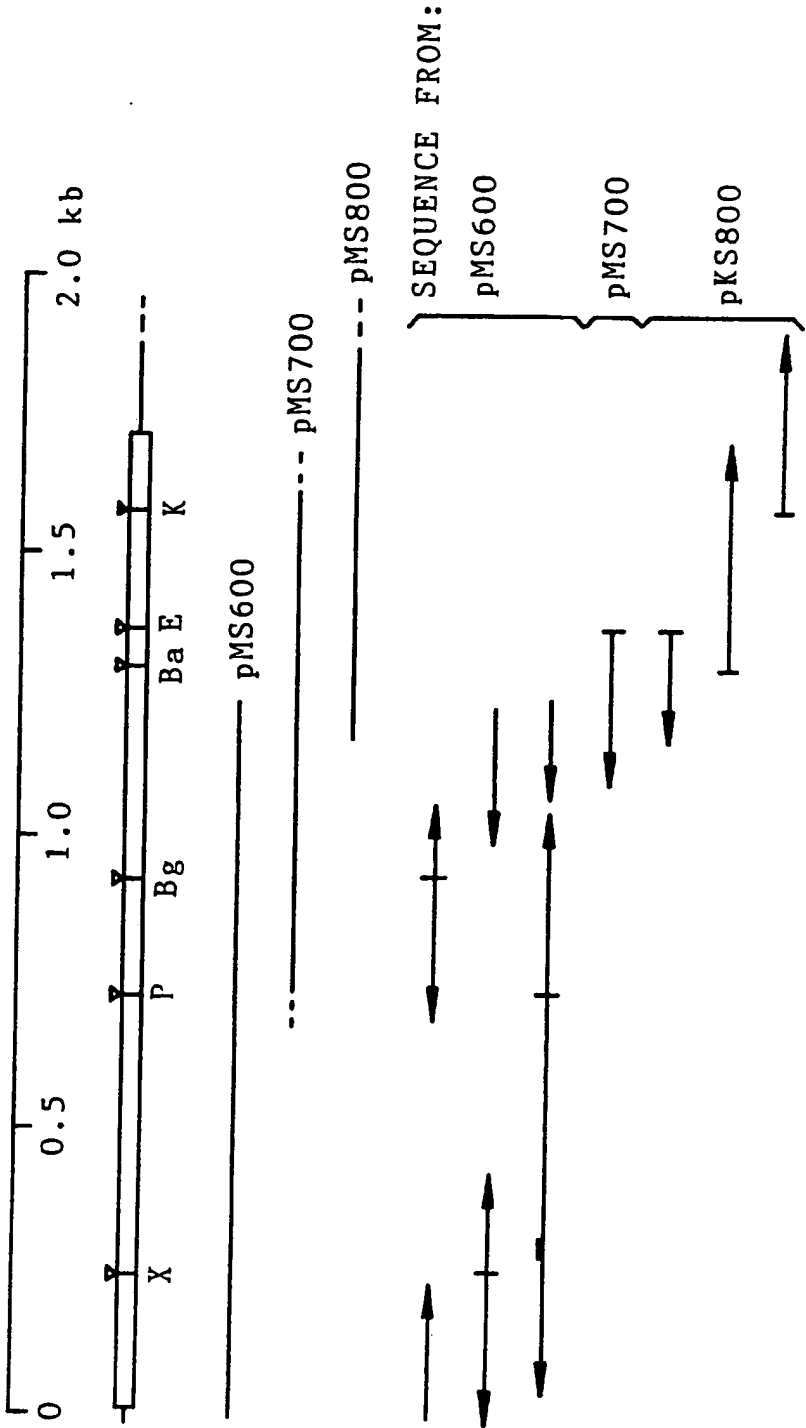
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CLAIMS

- 1
- 2
- 3 1. A 67kD protein of *Theobroma cacao*, or a fragment thereof.
- 4
- 5 2. A 47kD protein of *Th. cacao*, or a fragment thereof.
- 6
- 7 3. A 31kD protein of *Th. cacao*, or a fragment thereof.
- 8
- 9 4. A protein as claimed in claim 1, 2 or 3, having at least part of the
- 10 sequence shown in Figure 2.
- 11
- 12 5. A fragment as claimed in any one of claims 1 to 4, which comprises at
- 13 least four amino acids.
- 14
- 15 6. A protein or fragment as claimed in any one of claims 1 to 6, which is
- 16 recombinant.
- 17
- 18 7. Recombinant or isolated nucleic acid coding for a protein or fragment as
- 19 claimed in any one of claims 1 to 5.
- 20
- 21 8. Nucleic acid as claimed in claim 7 which is DNA.
- 22
- 23 9. Nucleic acid as claimed in claim 8, having at least part of the sequence
- 24 shown in Figure 2.
- 25
- 26 10. Nucleic acid as claimed in claim 7, 8 or 9, which is in the form of a
- 27 vector.
- 28
- 29 11. Nucleic acid as claimed in claim 10, wherein the vector is an expression
- 30 vector and the protein- or fragment-coding sequence is operably linked to a
- 31 promoter.
- 32
- 33

- 1 12. Nucleic acid as claimed in claim 11, wherein the expression vector is a
2 yeast expression vector and the promoter is a yeast pyruvate kinase (PK)
3 promoter.
4
- 5 13. Nucleic acid as claimed in claim 11, wherein the expression vector is a
6 bacterial expression vector and the promoter is a strong lambda promoter.
7
- 8 14. Nucleic acid as claimed in claim 11, 12 or 13, comprising a signal
9 sequence.
10
- 11 15. A host cell comprising nucleic acid as claimed in any one of claims 10 to
12 14.
13
- 14 16. A host cell as claimed in claim 15 which is *Saccharomyces cerevisiae*.
15
- 16 17. A host cell as claimed in claim 15 which is *E. coli*.
17
- 18 18. A process for the preparation of a protein or fragment as claimed in any
19 one of claims 1 to 5, the process comprising coupling successive amino acids by
20 peptide bond formation.
21
- 22 19. A process for the preparation of a protein or fragment as claimed in any
23 one of claims 1 to 5, the process comprising culturing a host cell as claimed in
24 claim 15, 16 or 17.
25
- 26 20. A process for the preparation of nucleic acid as claimed in any one of
27 claims 7 to 14, the process comprising coupling together successive nucleotides
28 and/or ligating oligo- or poly-nucleotides.
29
30
31
32
33

FIG. 1



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M V I S K S P F I V L I F S L L
 AAGCATAGCAAAATATGGTGATCAGTAAGTCTCCTTTTCATAGTTTGTGATCTTCTCTTCT
 10 20 30 40 50 60

 L S F A L L C S G V S A Y G R K Q Y E R
 CCTTTCTTTTGGTGTGCTTTTGTCTGGTGTGAGCGCCCTATGGCAGAAACAATATGAGCG
 70 80 90 100 110 120

 D P R Q Q Y E Q C Q R R C E S E A T E E
 TGATCCTCGACAGCAATACGAGCAATGCCAGAGCGGATCGAGTCGGAAGCGACTGAAGA
 130 140 150 160 170 180

 R E Q E Q C E Q R C E R E Y K E Q Q R Q
 AAGGAGCAAGAGCAGTGTGAACAACGCTGTGAAGGGAGTACAAGGAGCAGCAGAGACA
 190 200 210 220 230 240

 Q E E E L Q R Q Y Q Q C Q G R C Q E Q Q
 GCAAGAAGAAGAGCTTCAAGGCAATACCAGCAATGTCAAGGGCGTTGTCAAGAGCAACA
 250 260 270 280 290 300

 Q G Q R E Q Q C Q R K C W E Q Y K E Q
 ACAGGGGCAGAGAGCAGCAGCAGTGGCCAGAGAAAATGCTGGGAGCAATATAAGGAACA
 310 320 330 340 350 360

 E R G E H E N Y H N H K K N R S E E E
 AGAGAGAGGCGAGCAGAGAATTACCATAATCACAAAAAATAGGAGCGGAAGAAGAAGA
 370 380 390 400 410 420

 G Q Q R N N P Y Y F P K R R S F Q T R F
 AGGGCAACAAAGAAACAATCCTTACTATTTCCTTAAAGAAGATCATTCCAAACGATT
 430 440 450 460 470 480

FIG. 2A

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R D E E G N F K I L Q R F A E N S P L
 CAGGGATGAAGAGGGCAACTTCAAGATCCTCCAGAGGTTTGCTGAGAACTCTCCTCCACT
 490 500 510 520 530 540

 K G I N D Y R L A M F E A N P N T F I L
 CAAGGGCATCAACGATTACCGCTTGCCATGTTGGAAGCAAATCCCAACACTTTTATTCT
 550 560 570 580 590 600

 P H H C D A E A I Y F V T N G K G T I T
 TCCGCACCACTGTGATGCTGAGGCAATTACTTCGTGACAAACGGAAGGGGACAATTAC
 610 620 630 640 650 660

 F V T H E N K E S Y N V Q R G T V V S V
 GTTGTGACTCATGAACAAAGAGTCCTATAATGTACAGCGTGGAACAGTAGTCAGCGT
 670 680 690 700 710 720

 P A G S T V Y V V S Q D N Q E K L T I A
 TCCTGCAGGAAGCACTGTTTACGTGGTTAGCCAAAGACCAACGAAGAGCTAACCATAGC
 730 740 750 760 770 780

 V L A L P V N S P G K Y E L F F P A G N
 TGTGCTCGCCCTGCTTAATTCTCCTGGCAAATATGAGTTATTCTTCCCGCTGGAAA
 790 800 810 820 830 840

 N K P E S Y Y G A F S Y E V L E T V F N
 TAATAAACCTGAATCATATTACGGAGCCCTTCAGCTATGAAGTTCTTGAGACCCGTCTTCAA
 850 860 870 880 890 900

 T Q R E K L E E I L E E Q R G Q K R Q Q
 TACACAAAGAGAGAAGCTGGAGGAGATCTTGGAGGAACAGAGAGGGCAGAGAGGCAGCA
 910 920 930 940 950 960

FIG. 2B

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G Q Q G M F R K A K P E Q I R A I S Q Q
 GGGCAGCAGGGTATGTTCCGGAAGCCAAAGCCAGAGCAGATAAGAGCAATAAGCCAACA
 970 980 990 1000 1010 1020

 A T S P R H R G G E R L A I N L L S Q S
 AGTACTTCTCCAAGGCACAGAGGGGGGAGAGACTTGCCATCAATCTATTGAGCCAATC
 1030 1040 1050 1060 1070 1080

 P V Y S N Q N G R F F E A C P E D F S Q
 GCCTGTCTACTCCAACCAAAACGGACGCTTCTTTGAGGCTTGTCCTGAGGACTTCAGTCA
 1090 1100 1110 1120 1130 1140

 F Q N M D V A V S A F K L N Q G A I F V
 ATTTCAAGAACATGGATGTCGCTGTTTCAGCCTTCAAACTGAATCAGGGAGCCATATTGT
 1150 1160 1170 1180 1190 1200

 P H Y N S K A T F V V F V T D G Y G Y A
 GCCACACTACAATTCTAAGGCTACATTCTGTTGTTGTCACGGACGGATATGGGTACGC
 1210 1220 1230 1240 1250 1260

 Q M A C P H L S R Q S Q G S Q S G R Q D
 TCAAATGGCTTGCCCGCATCTCTCCAGACAGAGCCAGGATCCCAAAGTGGAAGGCAAGA
 1270 1280 1290 1300 1310 1320

 R R E Q E E E S E E T F G E F Q Q V K
 CAGAAGAGAACAAGAAGAGTCAGAAGAGGAGACACATTTGGAGAATTCCAGCAGGTCAA
 1330 1340 1350 1360 1370 1380

 A P L S P G D V F V A P A G H A V T F F
 AGCCCCATTGTACCTGGTGACGCTTTGTAGCCCCGGCAGGCCATGCAGTTACATTCTT
 1390 1400 1410 1420 1430 1440

FIG. 2C

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A S K D Q P L N A V A F G L N A Q N N Q
 TGCATCCAAAGACCCCTGAATGCAGTTGCGTTTGGACTCAACGCCCAAGAACCA
 1450 1460 1470 1480 1490 1500

 R I F L A G K K N L V R Q M D S E A K E
 GAGAATTTCCCTTGCAGGGGAAAAAGAACTTGGTCAGACAAATGGATAGCGGCAAGGA
 1510 1520 1530 1540 1550 1560

 L S F G V P S K L V D N I F N N P D E S
 GTTATCATTTGGGTACCATCGAAATTGGTAGATAATATATTCAACAACCCGGATGATC
 1570 1580 1590 1600 1610 1620

 Y F M S F S Q Q R R Q R D E R R G N P L
 GTATTTCATGTCCTTCTCTCAACAGAGCGCGTCGAGATGAAAGGAGGGCAATCCCTT
 1630 1640 1650 1660 1670 1680

 A S I L D F A R L F *
 GGCCCTCAATTCTGGACTTTGCCCCGCTTGTCTTAAGCAGCTGCTTCCACTTTTGTATCAGA
 1690 1700 1710 1720 1730 1740

 CATGCAGAGGCATGTAATGCAATAAATAAGTTGGCCCTATGTAAAGAGGAGAGATTGCT
 1750 1760 1770 1780 1790 1800

 TTTGTCTTGTCTAACCTTGTTTTGAACCTAGTAAACTTTCAATGTAATGAGAGTTGTTAT
 1810 1820 1830 1840 1850 1860

 CTTTCTA

FIG. 2D

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10 MVISKSPFIV LIFSLLSFA LLCSGVSAYG RKQYERDPRQ QYEQCQRCE SEATEEREQE 60
 20
 30
 40
 50
 60
 70 QCEQRCEREY KEQQRQEEELQRQYQQCG RCQEQQGQR EQQCCQRKCW EQYKEQERGE 120
 80
 90
 100
 110
 120
 130 EEPGSQF ANPAYHF 180
 140
 150
 160
 170
 180
 HENYHNHKKR RSEEEGQQR NNPLYFFPKRR SFQTRFRDEE GNFKILQRF A ENSPPLKGIN
 190
 200
 210
 220
 230
 240
 DYRLAMFEAN PNTFILPHHC DAEAIYFVTN GKGITITFVTH ENKESYNVQR GTVVSVPAGS
 250
 260
 270
 280
 290
 300
 TVVVSQDNQ EKLTI AVLAL PVNSPGKYEL FFPAGNNKPE SYYGAFSVEV LETVFNTORE
 310
 320
 330
 340
 350
 360
 KLEEIIEEQR GQKRQQGQG MFRKAKPEQI RAISQQATSP RHRGGERLAI NLLSQSPVYS
 370
 380
 390
 400
 410
 420
 NQNGRFFEAC PEDFSQFQNM DVAVSFAFLN QGAIFVPHYN SKATFVVVFT DGYGYAQMAMC
 430
 440
 450
 460
 470
 480
 PHLSRQSQS QSGRQDRREQ EEESEETFG EFQQVKAPLS PGDVFVAPAG HAVTFFASKD
 490
 500
 510
 520
 530
 540
 QPLNAVAFGL NAQNNQRIFL AGKKNLVRQM DSEAKELSFG VPSKLVDNIF NNPDSEYFMS
 550
 560
 570
 580
 590
 600
 FSQQRQRDE RRGNPIASIL DFARLF

FIG.3

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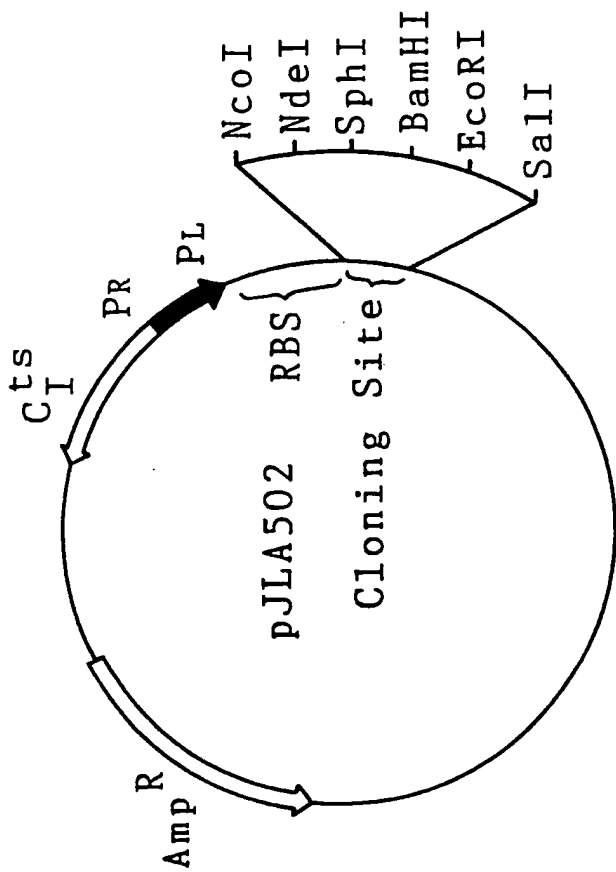
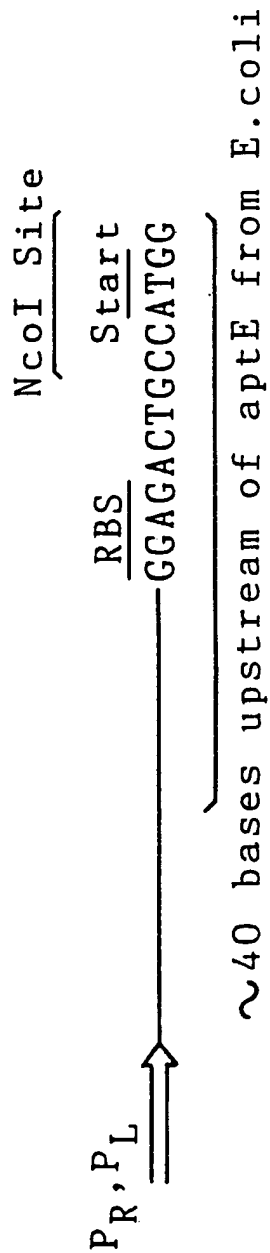
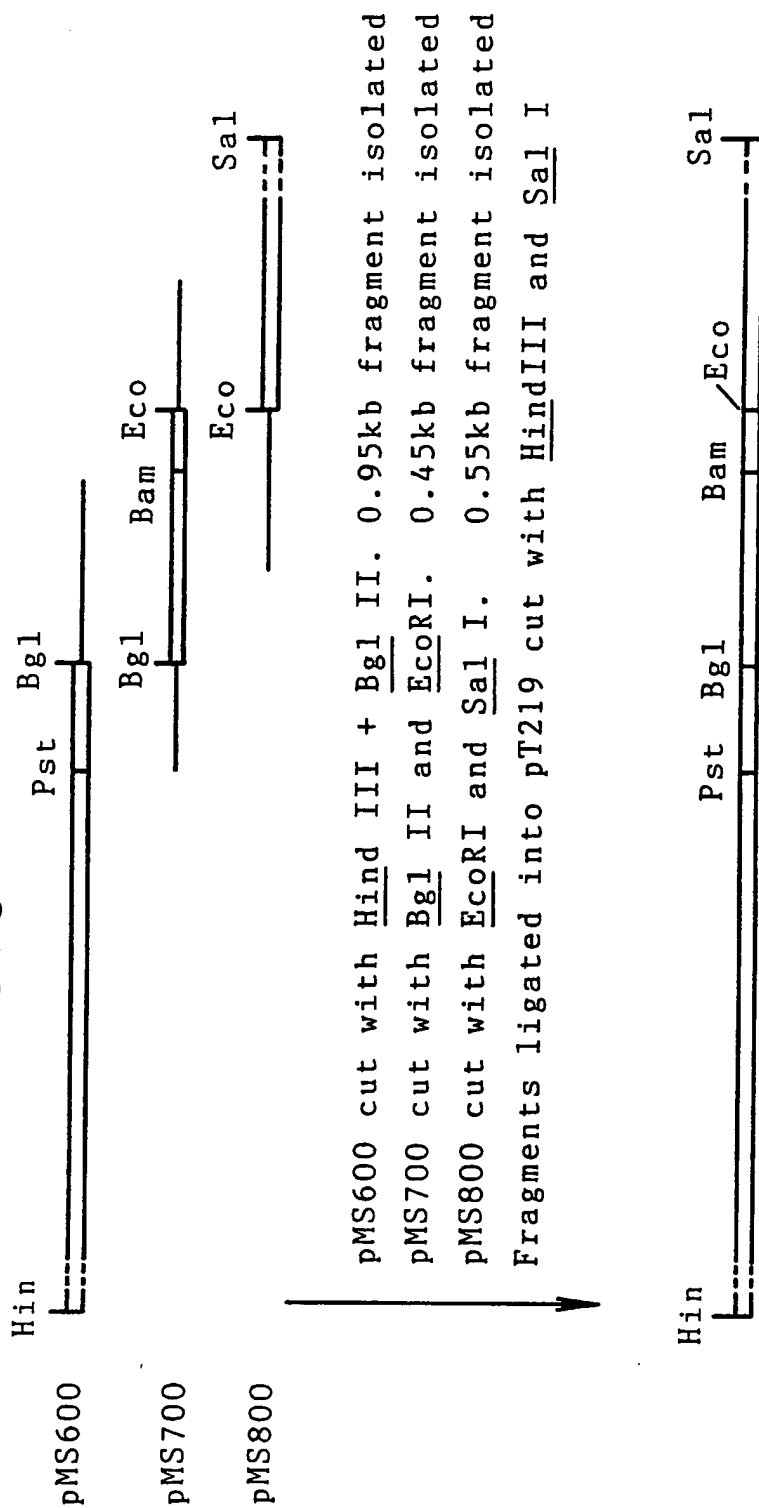


FIG. 5



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FIG. 6



key

key

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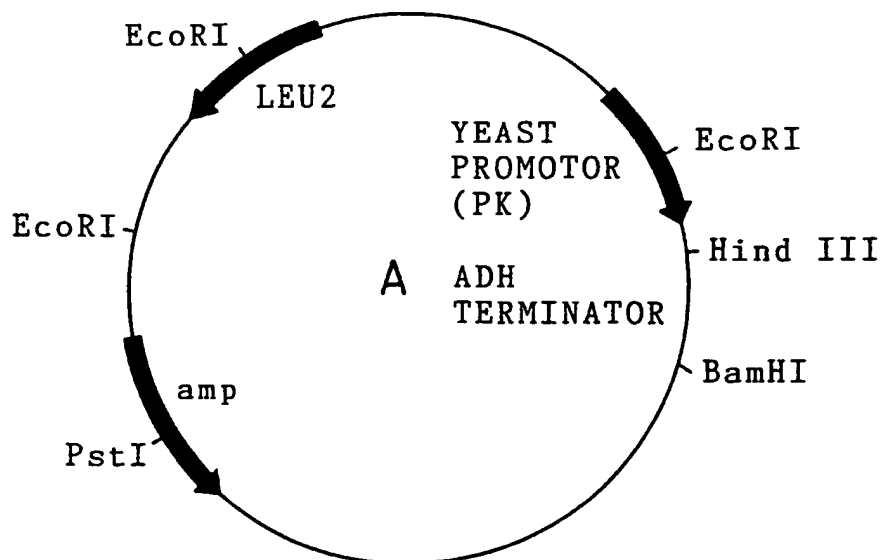


FIG. 7

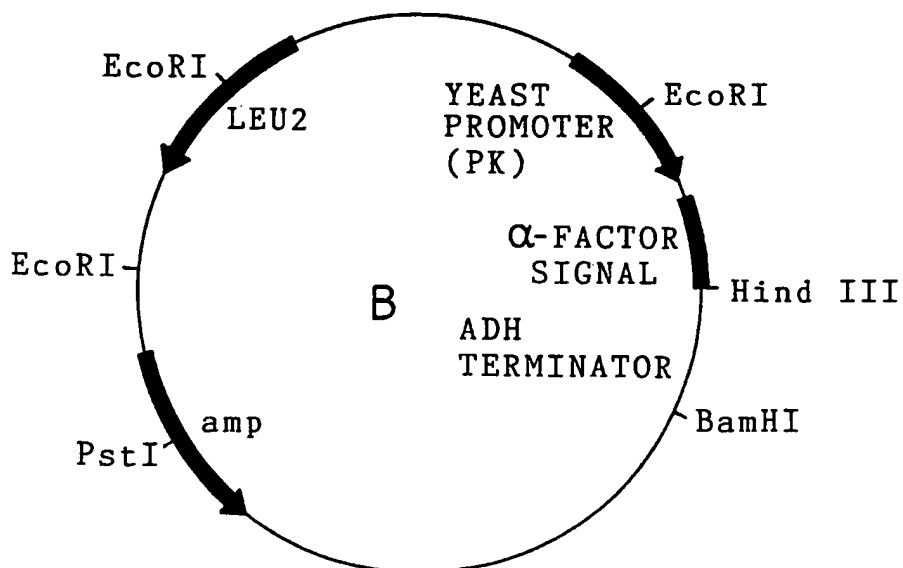


FIG. 8A

YEAST PYRUVATE
KINASE GENE

TTTACAAGACACCAATCAAACAATAAAACATCATCAATGTCTAGA
MetSerArg

SEQUENCE ALTERED
TO CREATE CLONING
SITE

TTTACAAGCTTCCAATCAAACAATAAAACATCATCAATGTCTAGA
Hin MetSerArg

HIN-NCO LINKERS

+

AGCTTCCAATCAAACAATAAAACATCATCAC
AGGTTAGTTTGTGTTATTTGTAGTAGGTAC

67KD EXPRESSION
VECTOR A

TTTACAAGCTTCCAATCAAACAATAAAACATCATCAATGGTGATC
Hin MetValIle
Nco

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FIG. 8B

1 231
 Met-----GluGlyValSerLeuAspLysArgGlu
 ATG-----GAAGGGGTAAGCTTGGATAAAAGAGAG

Hin

YEAST ALPHA-FACTOR
 SIGNAL SEQUENCE

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AGCTTGGATAAAAGAGC
 ACCTATTTTCTCGGTAC

HIN-NCO LINKERS

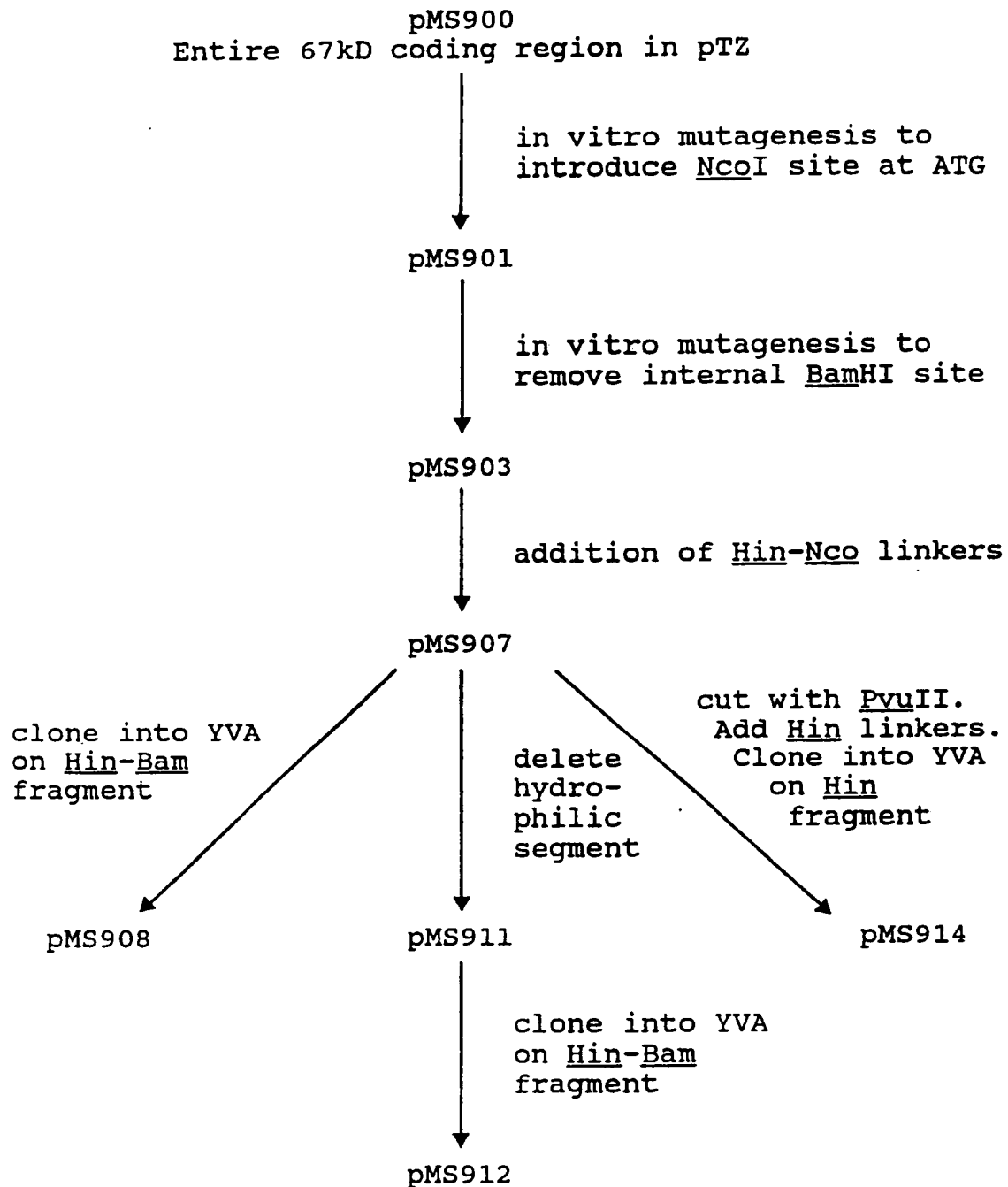
Met---GluGlyValSerLeuAspLysArgAlaMetAlaLeu
 ATG---GAAGGGGTAAGCTTGGATAAAAGAGCCCATGGCGTTG

Nco

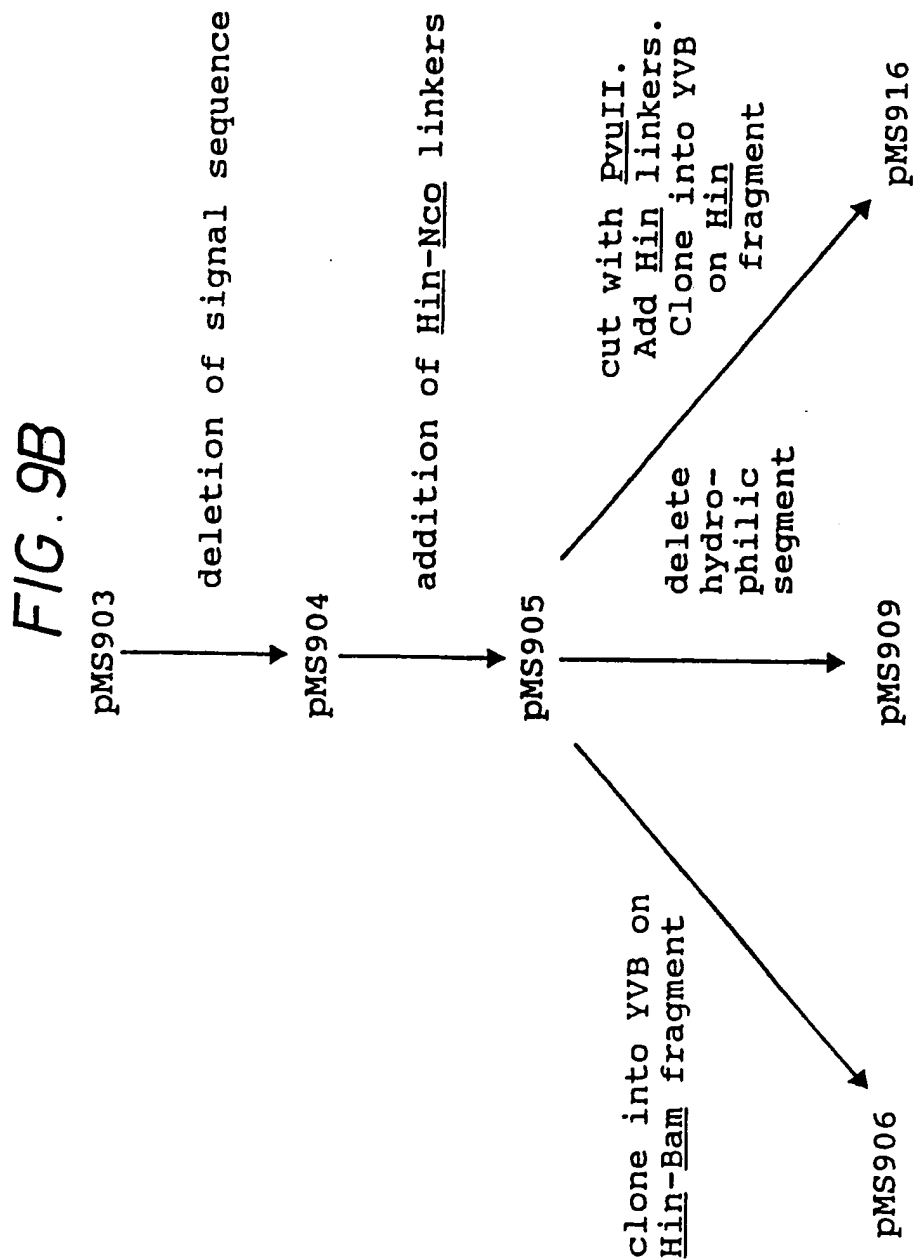
Hin

IN-PHASE FUSION OF
 67KD CODING REGION

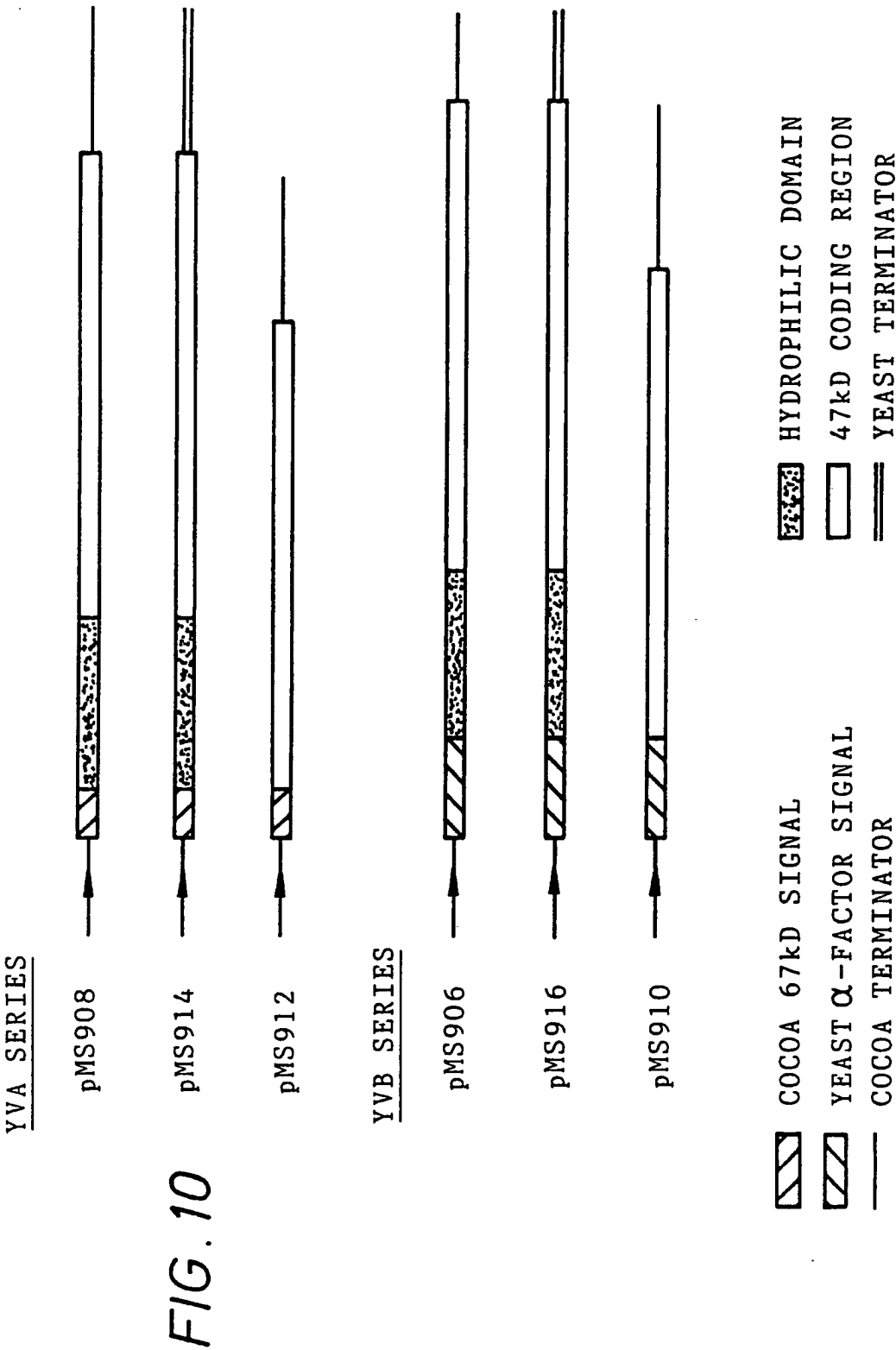
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FIG. 9A

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SUBSTITUTE SHEET



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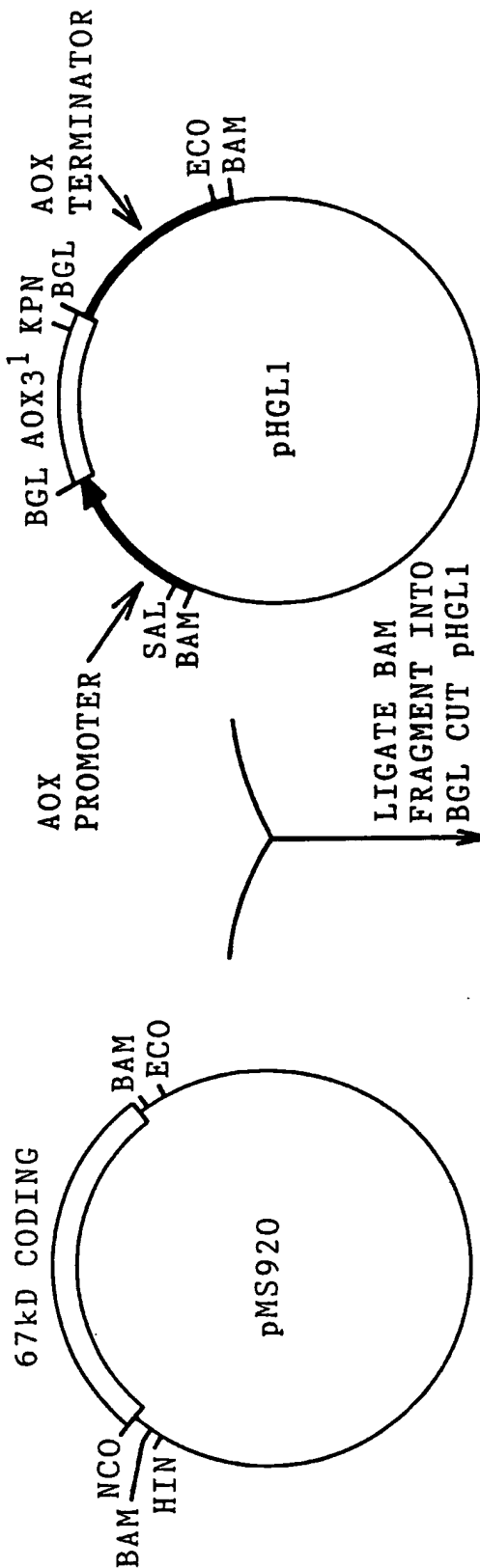
HIN NCO 67kD CODING HIN
HIN-HIN FRAGMENT
FROM pMS914

↓
FILL IN OVER HANGING ENDS.
↓
LIGATE ON BAM LINKERS.
↓
CUT WITH BAM

BAM NCO 67kD CODING BAM

↓
LIGATE INTO BAM CUT
PT2

FIG. 11A



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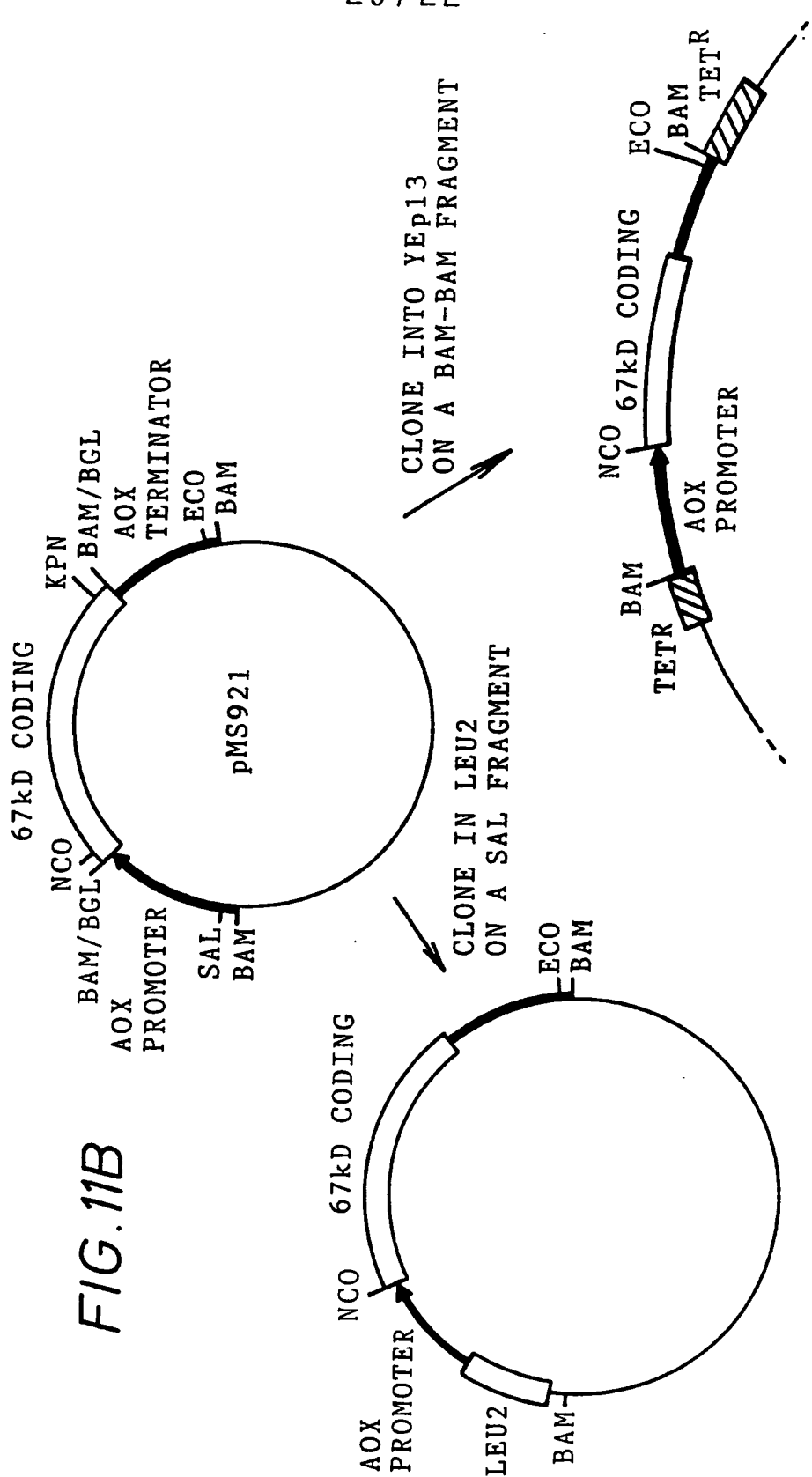


FIG.11B

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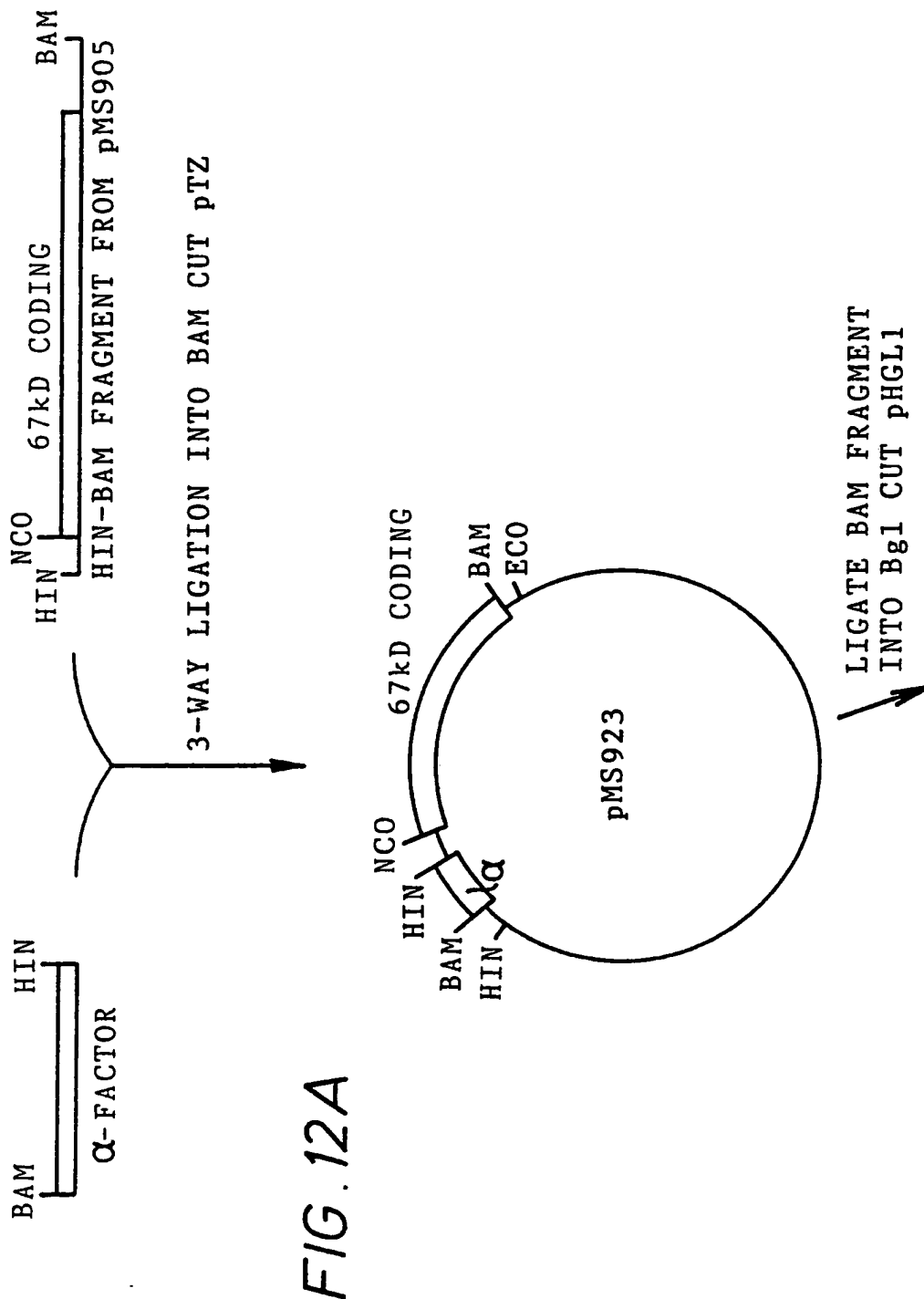
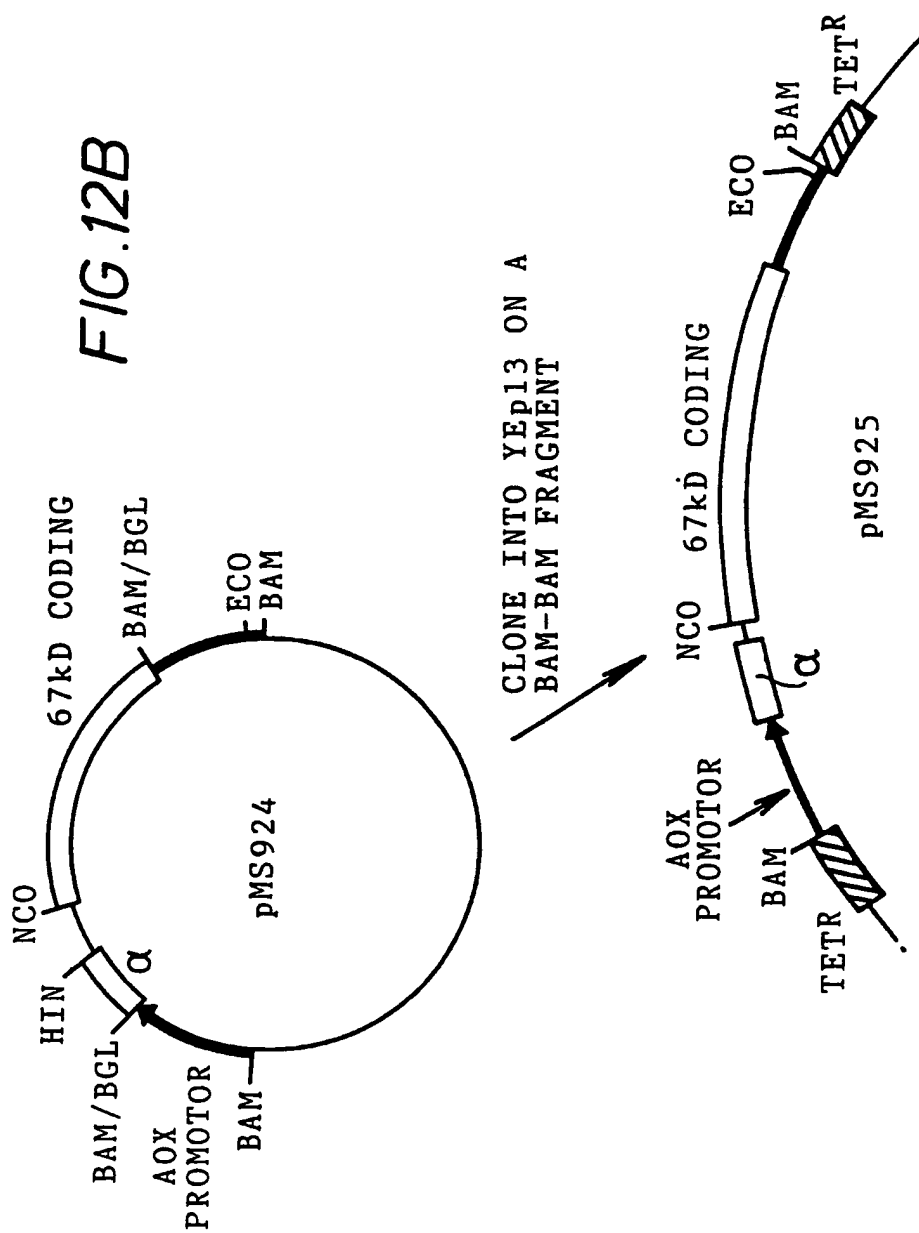


FIG. 12A

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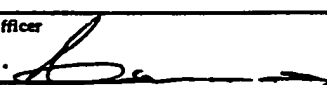
FIG. 12B



INTERNATIONAL SEARCH REPORT

International Application No

PCT/GB 91/00914

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ⁶		
According to International Patent Classification (IPC) or to both National Classification and IPC Int.Cl. 5 C12N15/29 ; C07K13/00 ; C12N1/21 ; C12N1/19		
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁷		
Classification System	Classification Symbols	
Int.Cl. 5	C07K ; C12N	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁸		
III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹		
Category ¹⁰	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
X	J. SCI. FOOD AGRIC. vol. 33, 1982, pages 1291 - 1304; BIEHL, B., ET AL: 'Vacuolar storage proteins of cocoa seeds and their degradation during germination and fermentation ' see the whole document ---	1-6
X	J. FOOD SCIENCE vol. 50, 1985, pages 946 - 950; FRITZ, P. J., ET AL: 'Cocoa seeds: Changes in protein and polysomal RNA during development ' see the whole document --- -/-	1-6
¹⁰ Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "A" document member of the same patent family		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
30 SEPTEMBER 1991	20. 11. 91	
International Searching Authority	Signature of Authorized Officer	
EUROPEAN PATENT OFFICE	MADDOX A.D. 	

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category °	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claims No.
X	CAFE CACAO THE vol. 34, no. 1, January 1990, pages 23 - 26; PETTIPHER G. L.: 'The extraction and partial purification of cocoa storage proteins '	1-6
Y	see the whole document	7-20
Y	ABSTR. PAP. AM. CHEM. SOC. vol. 188, 1984, BIOL 148 WILSON, M. R., ET. AL.: 'Cocoa theobroma -cacao seed complementary DNA library ' see the abstract 148	7-20
X	PLANT MOL. BIOL. vol. 9, no. 6, 1987, pages 533 - 546; CHLAN C. A., ET. AL.: 'Developmental biochemistry of cottonseed embryogenesis and germination. XIX. Sequences and genomic organization of the alpha-globulin, vicilin , genes of cottonseed. ' see figures 3,4	4-11,14, 15,17,20
X	CHEMICAL ABSTRACTS, vol. 109, no. 17, Columbus, Ohio, US; abstract no. 143880s, WATSON, MARTIN D. 'Isolation and expression of a pea vicilin cDNA in the yeast Saccharomyces cerevisiae ' see abstract	4-11, 14-16,19
X	PROC. NATL. ACAD. SCI. U. S. A. vol. 82, no. 2, January 1985, pages 334 - 338; CRAMER J H: 'Expression of phaseolin cDNA genes in yeast under control of natural plant DNA sequences. ' see the whole document	4-11, 14-16,19
X	CHEMICAL ABSTRACTS, vol. 106, no. 13, Columbus, Ohio, US; abstract no. 98280, CRAMER, JANE HARRIS 'Signal peptide specificity in posttranslational processing of the plant protein phaseolin in Saccharomyces cerevisiae ' see abstract	4-11, 14-16,19

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claims No.
X	PLANT MOL BIOL vol. 11, 1988, pages 683 - 695; HIGGINS T.J.V., ET.AL.: 'The sequence of a pea vicilin gene and its expression in transgenic tobacco plants ' see page 684 methods section ---	4-11, 15, 17, 19